In Vitro Variability Among Isolates of Eight Phytophthora Species in Response to Phosphorous Acid

M. D. Coffey and L. A. Bower

Associate professor and staff research associate, Department of Plant Pathology, University of California, Riverside 92521.

This research was funded in part by a grant from the California Avocado Commission. We also acknowledge the assistance given by Jean Paviot and Doug Johnson of Rhône-Poulenc in this project.

The authors thank Jaswinder Grover for technical work and BettyAnn Merrill for typing the manuscript. Accepted for publication 8 February 1984.

ABSTRACT

Coffey, M. D., and Bower, L. A. 1984. In vitro variability among isolates of eight *Phytophthora* species in response to phosphorous acid. Phytopathology 74:738-742.

The ED₅₀ values for inhibition of mycelial growth in vitro by phosphorous acid (H_3PO_3) ranged from 5.2 to 224.4 $\mu g/ml$ for nine *Phytophthora* species. Among the most sensitive species were *P. citricola*, *P. citrophthora*, and *P. cinnamoni*. One of the most tolerant species was *P. megasperma* f. sp. *medicaginis*. At the extreme of the range, *P. infestans* from potato had an ED₅₀ value of 224.4 $\mu g/ml$. Even among a group of similarly sensitive isolates, it was possible to differentiate them at a species

level in terms of their growth response to H_3PO_3 . Isolates of *P. citricola* from avocado were inhibited by 48.3–67.6%, and those of the A2 mating type of *P. cinnamomi* by 11.3–38.5%, in the presence of 5 μ g/ml H_3PO_3 . Isolates of *P. citrophthora* from citrus were inhibited 80.3–89.3%, compared with 27.9–58.8% for isolates of *P. parasitica* in the presence of 10 μ g/ml H_3PO_3 .

Additional key words: Aliette, efosite-Al, fosetyl-Al, fosetyl-Na, fungicide, phosethyl-Al.

The fungicide fosetyl-Al (aluminum tris-O-ethyl phosphonate), which is produced as an 80% wettable powder formulation by Rhône-Poulenc Agrochimie of Lyon, France, under the trade name of Aliette®, is active against many soilborne plant diseases caused by *Phytophthora* (1,6,17,23,24). Until recently, its mode of action in vivo was thought to involve primarily an alteration in host metabolism rendering the plant less susceptible to fungal attack (2,3,9,13-15,17,20,22,24). In part, this conclusion was reached because fosetyl-Al was found to have very low activity against mycelial growth of Phytophthora spp. in vitro (2-4, 10,18,19,21,22,24). In one study, however, it was observed that the ED₅₀ for linear mycelial growth of Phytophthora citrophthora (Smith & Smith) Leonian was only 56 µg/ml, although the equivalent ED₅₀ for *P. parasitica* Dastur was 929 μ g/ml (10). Some significant in vitro activity has been detected at different points in the life cycle of several species of Phytophthora (4,10,21). For instance, sporangium formation in both P. parasitica and P. citrophthora was completely inhibited by fosetyl-Al at only 10 μg/ml (10). Similarly, with P. cactorum (Lebert & Cohn) Schroeter and P. citricola Sawada, concentrations of fosetyl-Al at 10-40 $\mu g/ml$ were inhibitory to the formation of both sporangia and zoospores (4).

Recently a more critical examination of the effects of fosetyl-Al on mycelial growth of several *Phytophthora* species in vitro demonstrated that the phosphate content of the medium has an effect on activity (11). On a high-phosphate medium, fosetyl-Al caused only a slight inhibition of mycelial growth of two *Phytophthora* species. However, there was strong inhibition on a medium containing a low content of phosphate with ED 50 values for fosetyl-Al for several isolates of two *Phytophthora* species ranging from 45 to 54 μ g/ml. Presumably high levels of phosphate could interfere with the uptake of fosetyl-Al by the fungus (11).

In plant tissues, fosetyl-Al is thought to degrade to phosphorous acid (H_3PO_3) (3,20,22,23). In vitro, H_3PO_3 was found to be much more active than fosetyl-Al against mycelial growth of several *Phytophthora* species and the phosphate content of the medium

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. § 1734 solely to indicate this fact.

had little or no effect on efficacy (11). H₃PO₃ appears to be the toxophore responsible for inhibition of *Phytophthora* species when host plants are treated with fosetyl-Al (11).

The purpose of this present study was to establish the in vitro sensitivities of isolates of eight *Phytophthora* species to H_3PO_3 .

MATERIALS AND METHODS

In all the in vitro tests, H₃PO₃ and fosetyl-Na (sodium ethyl phosphonate) were added to agar media before autoclaving and the pH was adjusted to 6.2 with KOH. Fosetyl-Na was used in preference to fosetyl-Al since the latter causes a precipitation of aluminum hydroxide.

The solid media employed were either a modified Ribeiro's synthetic medium (16) containing 0.084 mM (with fosetyl-Na) or $0.84 \text{ mM KH}_2\text{PO}_4$ (with H_3PO_3) and no β -sitosterol (11), cornmeal agar, or rye-seed medium A (5). A 0.4-cm-diameter agar disk from an actively growing colony was used as the source of inoculum. Agar plates were incubated in the dark at 24 C, except those of *P. infestans* (Montagne) de Bary, which were incubated at 21 C. Radial linear growth was determined by measuring colony diameters at two positions on the fungal colony. Treatments were carried out in triplicate, and the experiments were repeated two times. Controls consisted of the same media without H_3PO_3 or fosetyl-Na. The levels of H_3PO_3 used were determined in preliminary experiments and selected because they allowed good differentiation of the various isolates being tested.

 ED_{50} values were calculated from linear regression lines obtained by plotting the percent inhibition of mycelial growth against the log concentration of H_3PO_3 . The standard errors of the ED_{50} values were calculated from a linear regression analysis of the H_3PO_3 dosage against the growth inhibition. The fungal isolates were selected from the collection of *Phytophthora* at the University of California, Riverside. These isolates had never been exposed to fosetyl-Al in the field.

RESULTS

A comparison of single isolates of *Phytophthora* demonstrated that the growth responses obtained with fosetyl-Na and H₃PO₃ were almost exactly parallel, although H₃PO₃ was much more inhibitory (Table 1). Consequently, further comparisons were carried out using H₃PO₃ only.

TABLE 1. Inhibition of radial growth of an isolate of four *Phytophthora* species on a modified Ribeiro's medium amended with either fosetyl-Na or phosphorous acid (H₃PO₃)^w

Species		Inhibition (%) ^x of radial growth by fosetyl-Na at ($\mu g/ml$):			ED ₅₀ values (μg/ml)	
	Isolate	80.5	241.0	482.0	Fosetyl-Na	H ₃ PO ₃ ^y
P. citricola	P1273	56.0 a²	79.4 a	92.2 a	82.0	7.0
P. cinnamomi	Pc402	24.5 b	52.7 b	76.4 b	202.8	11.9
P. capsici	P1319	23,6 b	37.5 c	62.9 c	310.5	34.7
P. parasitica	M134	21.5 b	43.0 c	59.5 c	314.1	30.9

[&]quot;The medium at pH 6.2 contained 0.084 mM KH₂PO₄, no β-sitosterol, and the growth was measured at 4 days.

TABLE 2. In vitro responses to phosphorous acid (H₃PO₃) among isolates of *Phytophthora cinnamomi* and *P. citricola* from avocado and other hosts'

Phytophthora species and	Isolate Host		Inhibition $(\%)^y$ of radial growth by H_3PO_3 at $5 \mu g/m$	
mating type	Isolate	11080		
P. citricola	P475	Avocado	67.6 a²	
	P1287	Avocado	64.8 a	
	P1273	Avocado	57.3 b	
	P602	Avocado	53.8 b	
	P1277	Avocado	48.3 c	
P. cinnamomi A2	Pc290	Juniperus	44.8 c	
A2	Pc352	Avocado	38.5 d	
A2	Pc289	Cedrus	30.4 e	
A1	Pc300	Camellia	25.4 f	
A2	Pc356	Avocado	22.6 fg	
A2	Pc311	Avocado	22.5 fg	
A1	Pc21	Camellia	18.8 g	
A2	Pc336	Avocado	18.8 g	
A2	Pc402	Avocado	11.3 h	
A1	Pc271	Banksia	6.3 i	
Al	Pc97	Camellia	0.0 j	
A1	Pc138	Avocado	0.0 j	

^{*}The medium at pH 6.2 contained 0.84 mM KH₂PO₄, no β -sitosterol, and the growth was measured at 7 days.

TABLE 3. In vitro responses to phosphorous acid (H_3PO_3) among isolates of *Phytophthora citrophthora* and *P. parasitica* grown on a modified Ribeiro's medium^x

Phytophthora		Hoot	Inhibition (%) ^y of radial growth		
species	Isolate	Host	by H ₃ PO ₃ at 10 μg/ml		
P. citrophthora	M143	Citrus	89.3 a²		
•	P1163	Citrus	87.1 ab		
	M132	Citrus	85.1 ab		
	M117	Citrus	84.9 ab		
	M142	Citrus	84.1 ab		
	P776	Cacao	82.0 ab		
	M131	Citrus	80.3 b		
P. parasitica	T131	Citrus	58.8 c		
•	M114	Citrus	41.5 d		
	M141	Citrus	30.8 e		
	M134	Citrus	30.6 e		
	M152	Citrus	27.9 e		

^xThe medium at pH 6.2 contained 0.84 mM $\c KH_2PO_4$, no $\c \beta$ -sitosterol, and the growth was measured at 7 days.

TABLE 4. In vitro responses to phosphorous acid (H_3PO_3) among isolates of *Phytophthora megasperma* from different hosts grown on rye-seed medium^u

			Inhibition (%) ^v of radial growth by H ₃ PO ₃ at:	
Isolate	Host	Origin	20 μg/ml	50 μg/ml
P405	Soybean	Mississippi	61.9 a ^z	82.6 a ^z
P1139	Soybean	Wisconsin	53.1 b	79.0 a
P1258	Ficus	New Guinea	45.0 c	53.8 b
P1057	Alfalfa	California	34.7 d	51.3 b
D1-304*	Douglas-fir (D1)	Oregon	26.7 e	36.7 c
D1-306 ^w	Douglas-fir (D1)	Oregon	26.3 e	33.8 c
D2-C17*	Douglas-fir (D2)	Oregon	15.0 f	13.3 ef
AL2-508 (2A)	Alfalfa (AL2)	Oregon	11.8 fg	11.8 f
P1253	Chick-pea	Australia	6.3 gh	14.8 ef
P844	Alfalfa	California	2.2 hi	23.2 d
AL2-509 (3B) ^y	Alfalfa (AL2)	Oregon	0.0 hi	0.0 g
P147	Sugar cane	Louisiana	−1.5 i	20.3 de

^uRye-seed medium A, pH 6.2 (5).

TABLE 5. In vitro responses to phosphorous acid (H₃PO₃) among isolates of *Phytophthora megasperma* f. sp. *glycinea* and f. sp. *medicaginis* from alfalfa, soybean, and chick-pea^x

Isolate	Hosţ	Origin	Inhibition (%) ⁹ of radial growth by H ₃ PO ₃ at 20 µg/ml
P1139	Soybean	Wisconsin	61.8 a ^z
P509	Soybean	Mississippi	45.9 Ь
P406	Soybean	Mississippi	45.3 b
P506	Soybean	Mississippi	38.5 c
P510	Soybean	Mississippi	37.2 c
P405	Soybean	Mississippi	27.9 d
P1057	Alfalfa	California	27.9 d
P1253	Chick-pea	Australia	14.9 e
1133-2	Alfalfa	California	12.1 e
P1316	Alfalfa	California	9.1 ef
1129-4	Alfalfa	California	7.1 f
P127	Alfalfa	Australia	-13.6 g

^{*}Isolates were grown at 24 C on rye-seed medium A at pH 6.2 (5).

^xInhibition based on comparison to unamended medium.

^y The ED₅₀ values for H₃PO₃ are taken from Table 8.

² Means with the same letter are not significantly different (P = 0.05) according to Duncan's new multiple range test.

y Inhibition based on comparison to unamended medium.

²Means with the same letter are not significantly different (P=0.05) according to Duncan's new multiple range test.

y Inhibition based on comparison to unamended medium.

² Means with the same letter are not significantly different (P = 0.05) according to Duncan's new multiple range test.

The growth was measured at 5 days. Inhibition based on comparison to unamended medium.

[&]quot;Douglas-fir group one isolate.

^{*}Douglas-fir group two isolates.

y Alfalfa group two isolates according to Hansen and Hamm (13).

² Means with the same letter are not significantly different (P = 0.05) according to Duncan's new multiple range test.

y Inhibition based on comparison to unamended medium.

² Means with the same letter are not significantly different (P = 0.05) according to Duncan's new multiple range test.

The growth inhibition of 12 isolates, including six from avocado (*Persea americana* Mill.), of *P. cinnamomi* Rands A1 and A2 mating types (25) ranged from 0 to 44.8% with 5 μ g/ml H₃PO₃ compared to unamended media (Table 2). Five isolates of *P. citricola* from avocado were more strongly inhibited (Table 2).

Isolates of *P. parasitica* and *P. citrophthora* from citrus were compared for mycelial growth responses to H₃PO₃. *P. citrophthora* had a narrow range of inhibition from 80.3 to 89.3%, whereas isolates of *P. parasitica* were less inhibited (27.9–58.8%) (Table 3).

A definite pattern of responses emerged among a limited number of isolates considered representative of different groups of P. megasperma (Drechs.) (Table 4). Two soybean isolates (P. megasperma Drechs. f. sp. glycinea Kuan & Erwin) were the most sensitive to H_3PO_3 at $20 \mu g/ml$. The two isolates from Douglas-fir (13) responded similarly. Two isolates of P. megasperma Drechs. f. sp. medicaginis Kuan & Erwin, one from chick-pea (P1253), and the other from alfalfa (P844) also demonstrated similar tolerance to H_3PO_3 at $20 \mu g/ml$ (Table 4).

A closer examination of in vitro responses of a larger number of isolates of *P. megasperma* from soybean (f. sp. *glycinea*) and alfalfa (f. sp. *medicaginis*) revealed a tendency toward separation of the types based on their host origin (Table 5), but in some instances their responses were similar.

A limited study of isolates of *Phytophthora* spp. from cacao (*Theobroma cacao* L.) showed some differential responses. *P. palmivora* (Butler) Butler isolates were usually most sensitive to H₃PO₃, *P. capsici* (Leonian) isolates were much more tolerant, and *P. citrophthora* was intermediate in sensitivity (Table 6). There was, however, some overlap among the three species, with an

TABLE 6. In vitro responses to phosphorous acid (H₃PO₃) among isolates of *Phytophthora* species from cacao

Species	Isolate	Origin	Inhibition (%) ^y of radial growth by H_3PO_3 at $10 \mu g/ml$
P. palmivora	P922	Malaysia	81.4 a ^z
P. palmivora	P832	Trinidad	78.5 b
P. palmivora	P1020	Nigeria	73.6 c
P. citrophthora	P1213	Brazil	56.7 d
P. citrophthora	P1212	Brazil	56.1 d
P. palmivora	P736	Ghana	53.0 e
P. citrophthora	P776	Brazil	43.6 f
P. citrophthora	P1201	Brazil	40.1 g
P. capsici	P782	Cameroun	30.9 h
P. capsici	P632	Brazil	20.5 i
P. capsici	P1195	Mexico	-0.85 j

^xIsolates were grown on Difco cornmeal agar at 24 C.

TABLE 7. In vitro responses to phosphorous acid (H₃PO₃) among A1 mating type isolates of *Phytophthora infestans*^x

Isolate	Origin	Inhibition (%) ^y of radial growth by H ₃ PO ₃ at 200 μg/ml		
P1297	Ireland	71.2 a²		
P4	Mexico	69.0 ab		
P1296	Ireland	67.1 ab		
P1298	Wales	63.5 abc		
M3	Mexico	61.0 abc		
P1293	Scotland	57.6 bc		
M4	Mexico	57.1 bc		
P1300	Wales	52.9 c		
P1295	Scotland	40.3 d		
65	Mexico	30.4 d		

^xIsolates were grown on rye-seed medium A at 21 C (5).

isolate of *P. palmivora* (P736) responding similarly to *P. citrophthora* (Table 6).

All ten Al isolates of *P. infestans* tested were extremely insensitive to H₃PO₃ compared to isolates of the other *Phytophthora* species tested. At least 200 µg of H₃PO₃ per milliliter was required to obtain any significant growth inhibition of *P. infestans* (Table 7).

The ED₅₀ values for isolates of nine *Phytophthora* species from different hosts were calculated (Table 8) from their dosage-response curves (Fig. 1). Values for isolates grown on the synthetic medium ranged from a low of 5.2 μ g/ml for an isolate of *P. citrophthora* to 91.2 μ g/ml for a chick-pea isolate of *P. megasperma* f. sp. *medicaginis* (Table 8). On rye-seed medium, the ED₅₀ for an isolate of *P. megasperma* f. sp. *medicaginis* (P1316) was 88.9 μ g/ml, while an isolate of *P. infestans* had an ED₅₀ value of 224.4 μ g/ml (Table 8).

Slopes of the dosage-response curves (Fig. 1) indicate that two isolates of P. citricola were the most sensitive to H_3PO_3 , closely followed by two isolates of P. citrophthora from citrus and by two isolates of P. cinnamomi from avocado. Among the most insensitive isolates were a P. cactorum isolated from Rhaphiolepis indica, a P. megasperma from chick-pea, and isolate P1257 of P. boehmeriae.

DISCUSSION

Fosetyl-Al has been shown to provide excellent control of a number of soilborne plant diseases caused by *Phytophthora* (1,6,23,24), because foliar applications provide systemic protection through its strong basipetal mobility (1,17,24). However, this fungicide has not controlled potato late blight caused by *P. infestans* (1,17,23).

Fosetyl-Al was believed to have little in vitro activity against *Phytophthora* because ED₅₀ values frequently were found to be 500–1,000 μ g/ml or even greater (2,10,17–19,23,24). Recently, much greater activity was demonstrated by using a synthetic medium that was low in phosphate (0.084 mM KH₂PO₄); ED₅₀ values of 45–54 μ g/ml were obtained for several isolates of two *Phytophthora* species (11). In addition, it was determined that H₃PO₃ was 6–14 times more active than fosetyl-Al in inhibiting mycelial growth.

The present study established closely parallel effects of fosetyl-Na and H₃PO₃ on mycelial growth of four isolates of *Phytophthora*. It also confirmed the greater sensitivity of *Phytophthora* spp. to H₃PO₃ compared to fosetyl-Na.

TABLE 8. ED₅₀ values³ for inhibition of radial growth expressed in micrograms per milliliter for individual isolates of nine *Phytophthora* species varying in response to phosphorous acid (H₃PO₃)

Species	Isolate	ED_{50} $(\mu\mathrm{g/ml})$	Standard deviation ^b
P. citrophthora	M143	5.2	± 3.7
P. citricola	P1287	6.8	± 1.1
P. citricola	P1273	7.0	± 1.7
P. cinnamomi Al	Pc97	9.0	± 0.9
P. cinnamomi A2	Pc356	9.9	± 1.6
P. citrophthora	P1163	10.4	± 2.7
P. cinnamomi A2	Pc402	11.9	± 0.7
P. capsici	P1091	18.5	± 0.9
P. megasperma f. sp. glycinea	P405	22.3	± 2.6
P. capsici	P1314	30.6	± 1.5
P. parasitica	M134	30.9	± 2.3
P. capsici	P1319	34.7	± 4.8
P. boehmeriae	P1257	40.6	± 8.3
P. cactorum	P1235	67.1	± 11.6
P. megasperma (chick-pea)	P1253	91.2	± 69.0
P. megasperma f. sp. medicaginis	P1316°	88.9	± 14.8
P. infestans	P1300°	224.4	± 10.8

^a Isolates of *Phytophthora* grown on modified Ribeiro's medium.

^yThe growth was measured at 4 days. Inhibition based on comparison to unamended medium.

² Means with the same letter are not significantly different (P = 0.05) according to Duncan's new multiple range test.

^yThe growth was measured at 7 days. Inhibition based on comparison to unamended medium.

⁷ Means with the same letter are not significantly different (P = 0.05) according to Duncan's new multiple range test.

^bMean ± standard deviation of the mean based on a linear regression of the response (percent mycelial inhibition) plotted against the dosage (log concentration H₃PO₃).

These isolates were grown on cleared rye-seed medium.

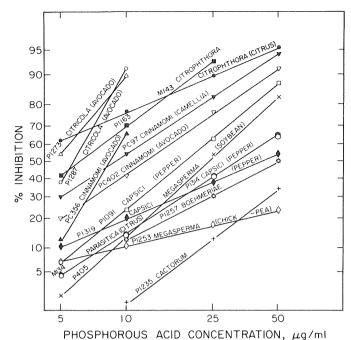


Fig. 1. Dosage-responses for 15 isolates of eight *Phytophthora* species, relating their growth response, expressed as a linear regression, to log concentration of phosphorous acid (H_3PO_3) .

When groups of isolates of different *Phytophthora* species were compared, responses to H_3PO_3 in many instances were found to be characteristic for a species. For instance, all isolates of *P. citricola*, *P. citrophthora*, and *P. cinnamomi* were found to be extremely sensitive with ED_{50} values of $5-10~\mu g/ml$. Despite their similar sensitivity, it was usually possible to separate *P. citricola* from *P. cinnamomi* by using a low level of H_3PO_3 (5 $\mu g/ml$).

Farih et al (10) compared single isolates of P. citrophthora and P. parasitica from citrus and obtained ED_{50} values of 56 and 929 $\mu g/ml$, respectively, for inhibition of mycelial growth by fosetyl-Al on V-8 juice agar. In the present study, several isolates of the two citrus pathogens demonstrated parallel, though more dramatic, responses to H_3PO_3 with ED_{50} values of 5–10 $\mu g/ml$ for P. citrophthora and about 30 $\mu g/ml$ for P. parasitica.

Results obtained with isolates of P. megasperma also produced some evidence for characteristic responses to H_3PO_3 based on formae speciales or morphological types (13). In particular, the isolates of P. megasperma f. sp. medicaginis (alfalfa host) were much less sensitive to H_3PO_3 , with ED_{50} values around 90 μ g/ml, compared with values around 20 μ g/ml for isolates of P. megasperma f. sp. glycinea (soybean host).

Similarly, among three species from cacao, *P. palmivora* was most sensitive, *P. citrophthora* was intermediate, and *P. capsici* was most tolerant to H₃PO₃, although there were some exceptions to this generalization. In view of the present gaps in our taxonomic knowledge of the "*P. palmivora* complex" (25), exceptions to the rule might be expected.

Metalaxyl, a fungicide that is also highly active against *Phytophthora* (17,25), also affects species differently in vitro (7). The specific mode of action of metalaxyl involves inhibition of an α -amanitin-insensitive, RNA polymerase-template complex (8). Interestingly, some of the most metalaxyl-sensitive species, such as *P. boehmeriae*, *P. megasperma* f. sp. *medicaginis*, and *P. parasitica* (7), were among the most tolerant to H₃PO₃. Conversely, species like *P. citrophthora* and *P. citricola*, which were relatively insensitive in vitro to metalaxyl (7), were very sensitive to H₃PO₃.

The high biological activity of H₃PO₃ against *Phytophthora* spp. indicates a specific effect of this toxophore against those pathogens. Such a concept must be reconciled with the alternative evidence that fosetyl-Al, and presumably therefore H₃PO₃, causes a stimulation of a host defense response (2,3,9,12,14,20–22). Further data will be required on the biological and chemical effect

of H_3PO_3 and fosetyl-Al, in vivo and in vitro, before this question can be resolved.

LITERATURE CITED

- 1. Bertrand, A., Ducret, J., Debourge, J. C., and Horrière, D. 1977. Étude des propriétés d'une nouvelle famille de fongicides: les monoéthylphosphites métalliques. Caractéristiques physico-chimiques et propriétés biologiques. Phytiatrie-Phytopharmacie 26:3-17.
- 2. Bompeix, G., Fettouche, F., and Saindrenan, P. 1981. Mode d'action du phoséthyl Al. Phytratrie-Phytopharmacie 30:257-272.
- 3. Bompeix, G., Ravisé, A., Raynal, G., Fettouche, F., and Durand, M. C. 1980. Modalités de l'obtention des nécroses bloquantes sur feuilles détachées de Tomate par l'action du tris-O-éthyl phosphonate d'aluminium (phoséthyl d'aluminium), hypothéses sur son mode d'action in vivo. Ann. Phytopathol. 12(4):337-351.
- Buchenauer, H., and Dercks, W. 1983. Mode of action of aluminium fosetyl in Peronosporales. (Abstr.) Page 75 in: Proc. Fourth Int. Congress of Plant Pathology, Melbourne, Australia.
- Caten, C. E., and Jinks, J. L. 1968. Spontaneous variability of single isolates of *Phytophthora infestans* 1. Cultural variation. Can. J. Bot. 46:329-348.
- 6. Clerjeau, M., and Beyriès, A. 1977. Étude comparée de l'action préventive et du pouvoir systémique de quelques fongicides nouveaux (phosphites-prothiocarbe, pyroxychlore) sur poivron vis-á-vis de *Phytophthora capsici* (Léon). Phytiatrie-Phytopharmacie 26:73-83.
- Coffey, M. D., and Bower, L. A. 1984. In vitro variability among isolates of six *Phytophthora* species in response to metalaxyl. Phytopathology 74:(In Press).
- 8. Davidse, L. C., Hofman, A. E., and Velthuis, G. C. M. 1983. Specific interference of metalaxyl with endogeneous RNA polymerase activity in isolated nuclei from *Phytophthora megasperma* f. sp. *medicaginis*. Exp. Mycol. 7:344-361.
- 9. Durand, M. C., and Sallé, G. 1981. Effet du tris-O-éthyl phosphonate d'aluminium sur le couple *Lycopersicon esculentum* Mill.—

 Phytophthora capsici Leon. Etude cytologique et cytochimique. Agronomie 1(9):723-731.
- Farih, A., Tsao, P. H., and Menge, J. A. 1981. Fungitoxic activity of efosite aluminum on growth, sporulation, and germination of Phytophthora parasitica and P. citrophthora. Phytopathology 71:934-936.
- Fenn, M. E., and Coffey, M. D. 1984. Studies on the in vitro and in vivo antifungal activity of fosetyl-Al and phosphorous acid. Phytopathology 74:606-611.
- Fettouche, F., Ravisé, A., and Bompeix, G. 1981. Suppression de la résistance induite-phoséthyl Al-chez la tomate à *Phytophthora capsici* avec deux inhibiteurs—Glyphosate et acide α-amino-oxyacétique. Agronomie 9:826.
- Hansen, E. M., and Hamm, P. B. 1983. Morphological differentiation of host-specialized groups of *Phytophthora megasperma*. Phytopathology 73:129-134.
- Langcake, P. 1981. Alternative chemical agents for controlling plant disease. Philos. Trans., R. Soc. Lond. B., Biol. Sci. 295:83-101.
- 15. Raynal, G., Ravisé, A., and Bompeix, G. 1980. Action du tris-O-éthyl phosphonate d'aluminium (phoséthyl d'aluminium) sur la pathogénie de *Plasmopara viticola* et sur la stimulation des réactions de défense de la vigne. Ann. Phytopathol. 12(3):163-175.
- 16. Ribeiro, O. K., Erwin, D. C., and Zentmyer, G. A. 1975. An improved synthetic medium for oospore production and germination of several *Phytophthora* species. Mycologia 67:1012-1019.
- 17. Schwinn, F. J. 1983. New developments in chemical control of *Phytophthora*. Pages 327-334 in: *Phytophthora*: Its Biology, Taxonomy, Ecology, and Pathology. D. C. Erwin, S. Bartnicki-Garcia, and P. H. Tsao, eds. Am. Phytopathol. Soc., St. Paul, MN.
- Smith, P. M. 1979. A study of the effects of fungitoxic compounds on *Phytophthora cinnamomi* in water. Ann. Appl. Biol. 93:149-157.
- Tey, C. C., and Wood, R. K. S. 1983. Effects of various fungicides in vitro on *Phytophthora palmivora* from cocoa. Trans. Br. Mycol. Soc. 80(2):271-282.
- Trique, B., Ravisé, A., and Bompeix, G. 1981. Modulation des infections à *Phytophthora* spp. provoquées chez la tomate. Agronomie 1(9):823-824.
- Vegh, I., Baillot, F., and Roy, J. 1977. Étude de l'activité de l'éthylphosphite d'aluminium (LS-74 783) vis-à-vis de *Phytophthora* cinnamomi Rands, agent due dépérissement des arbustes d'ornement. Phytiatrie-Phytopharmacie 26:85-95.
- 22. Vo, T.-H., Bompeix, G., and Ravisé, A. 1979. Rôle du tri-*O*-éthyl phosphonate d'aluminium dans la stimulation des réactions de défense des tissus de Tomate contre le *Phytophthora capsici*. C. R. Acad. Sci. Série D 288:1171-1174.

- 23. Williams, D. J., Beach, B. G. W., Horrière, D., and Marechal, G. 1977. LS 74-783, a new systemic fungicide with activity against phycomycete diseases. Pages 565-573 in: Proc. Br. Crop Prot. Conf., Pests and Diseases Section, 1977. 1045 pp.
- 24. Zentmyer, G. A. 1979. Effect of physical factors, host resistance and fungicides on root infection at the soil-root interface. Pages 315-328 in:
- The Soil-Root Interface. J. L. Harley and R. Scott-Russell, eds. Academic Press, London. 448 pp.
- 25. Zentmyer, G. A. 1983. The world of Phytophthora. Pages 1-7 in: Phytophthora: Its Biology, Taxonomy, Ecology, and Pathology. D. C. Erwin, S. Bartnicki-Garcia, and P. H. Tsao, eds. Am. Phytopathol. Soc., St. Paul, MN.